## MARGINATIN, A NEW GERMACRANOLIDE FROM VERNONIA SPECIES

## WILLIAM G. PADOLINA,\* NOBUJI NAKATANI,† HIROSUKI YOSHIOKA,‡ TOM J. MABRY§ and STEPHEN A. MONTI||

Department of Botany and | Department of Chemistry, University of Texas at Austin, TX 78712, U.S.A.

(Received 6 January 1974)

**Key Word Index**—*Vernonia marginata*; *V. fasciculata*; *V. arkansana*; Compositae; sesquiterpene lactone; germacranolide; marginatin.

**Abstract**—A new germacranolide, marginatin, has been isolated from three species of *Vernonia* from the central United States, *V. marginata*, *V. fasciculata* and *V. arkansana*, and its structure has been determined.

In the course of our biochemical systematic investigations of more than 45 species of *Vernonia* from throughout the Western Hemisphere, <sup>1-3</sup> a new germacranolide, which we have named marginatin (1), was found to be the major sesquiterpene lactone component of three species which occur in the central region of the United States: *V. marginata* (Torr.) Raf., range centered in West Texas; *V. fasciculata* Michx. and *V. arkansana*, DC, whose ranges are centered in the north central United States. Other closely related North American species were previously reported<sup>4</sup> to elaborate as their major sesquiterpene lactone constituent either glaucolide-A (2a) or B (2b); in some instances, marginatin occurs with (2a).

- \* Present address: Department of Chemistry, University of the Philippines at Los Banos' College, Laguna, Philippines.
- † Present address: Department of Agricultural Chemistry, The University of Tokyo, Bunkyo-ku, Tokyo, Japan.
- ‡ Present address: Takarazuka Research Laboratory, Sumitomo Chemical Co., Ltd., Takarazuka, Hyogo, Japan.
  - § To whom correspondence should be addressed.
- <sup>1</sup> PADOLINA, W. G. (1973) Ph.D. thesis, The University of Texas at Austin.
- <sup>2</sup> ABDEL-BASET, Z. H., SOUTHWICK, L., PADOLINA, W. G., YOSIHOKA, H., MABRY, T. J. and JONES, Jr., S. B. (1971) *Phytochemistry* **10**, 2201.
- <sup>3</sup> MABRY, T. J., ABDEL-BASET, Z. H., PADOLINA, W. G. and JONES, Jr., S. B. (1974) Biochem. Syst. and Ecol., prepared.
- <sup>4</sup> PADOLINA, W. G., YOSHIOKA, H., NAKATANI, N., MABRY, T. J., MONTI, S. A., DAVIS, R. E., COX, P. J., SIM, G. A., WATSON, W. H. and Wu, I. B., (1974) Tetrahedron 30, 1161.

Marginatin (1), which was first obtained from the chloroform extract of leaves of V. marginata, has the following properties:  $C_{22}H_{28}O_7$ , m.p.  $104-105^\circ$ ,  $[\alpha]_D^{25} - 6\cdot0^\circ$ , UV absorption at  $\lambda_{\text{max}} = 215$  nm ( $\epsilon$  25 600), IR 1760, 1710 and 1236 cm<sup>-1</sup>. The UV and IR data for marginatin indicated the presence of a conjugated γ-lactone ring. The PMR spectrum of marginatin (in acetone-d<sub>6</sub>; Table 1) exhibited a broad singlet at  $\delta$  4·92 (2H) and an acetate singlet at  $\delta$  2·02 (3H) which, by analogy with the PMR spectrum of glaucolide-A (2a) and B (2b), is indicative of an allylic acetate group. A triplet at  $\delta$  5·20 (1H, J 7 Hz) when shifted upfield by the addition of Pr(fod)<sub>3</sub> appeared as a double-doublet at  $\delta$  4·70 (1H, J 10, 4 Hz). Another doublet at  $\delta$  4·98 (1H, J 9 Hz) was also shifted upfield by Pr(fod)<sub>3</sub> and appeared as a broad doublet at  $\delta$  4·05 (1H, J 10 Hz) (0·02 mmol of Pr(fod)<sub>3</sub> was used to 0·19 mmol of 1). In the upfield region of the PMR spectrum of 1 a singlet appeared at 1·40 (3H) which could be assigned to a tertiary methyl group. In addition, a set of overlapping signals from  $\delta$  1·95 to 1·66 (9H) were assigned to three vinyl methyl groups. In the low-field region, a multiplet appeared at  $\delta$  7·00 (1H) which could be assigned to a  $\beta$ -proton of an  $\alpha$ -p-unsaturated ester.

SCHEME 1. CHEMICAL TRANSFORMATIONS OF MARGINATIN (1).

Catalytic hydrogenation of marginatin (1) over palladium-carbon in ethyl acetate yielded a mixture of the stereoisomers of tetrahydrodesacetoxymarginatin (3) (PMR spectral data in Table 1) in which hydrogenolysis of the  $C_{13}$  allylic acetate ester had occurred along with the reduction of two of the three double bonds in the compound. The conjugated  $\gamma$ -lactone moiety and an ester group were still present in (3) (IR 1750, 1725 cm<sup>-1</sup>). Furthermore, a new vinyl methyl signal at  $\delta$  2-03 (d, 3H, J 2 Hz) appeared in the PMR spectrum of (3) proving the presence of an allylic acetate moiety in marginatin which, by analogy with similar hydrogenolysis reactions for glaucolide-A and B<sup>1,4</sup> must be associated with the conjugated  $\gamma$ -lactone. The saturation of the other double bonds in (1) was indicated by the absence of the low-field vinyl proton signals in the PMR spectrum of (3). The functional groups in marginatin as indicated by the spectral and elemental analysis data were in accord with a germacranolide skeleton.

MS and PMR data indicated the presence of the tiglate ester in marginatin. The base peak in the MS of marginatin was at m/e 83 followed in intensity by another peak at m/e 55 (40%). Similarly, the base peak in the MS of 3 appeared at m/e 57 followed in intensity by another peak at m/e 85 (50%). These fragmentations are typical of the cleavage of esters of angelic, tiglic and senecioic acids. Both tiglic and angelic acids have a  $\beta$ -proton, cis to the carbonyl in the former and trans in the latter. Senecioic acid, however, only has an  $\alpha$ -proton. In the PMR spectrum of marginatin, a proton signal appeared at  $\delta$  7·00 (m, 1H), thus in the region where the  $\beta$ -proton of tiglic acid occurs. The remaining oxygen atom in marginatin could be assigned to an epoxide from consideration of the allylic alcohol (4) (and its acetate, 4b) obtained from the reaction of (3) in concd hydrochloric acid and methanol. A doublet at  $\delta$  4·52 (J 9 Hz) in the PMR spectrum of the alcohol (4) (in acetone- $d_6$ ; Table 1) could be assigned to  $H_5$  since it was shown to be coupled to  $H_6$  ( $\delta$  5·12, J 9, 2 Hz);  $H_6$  was also coupled to the  $C_{11}$  vinyl methyl group ( $\delta$  1·93, J 1 Hz). Since upon acetylation of (4a) to (4b) the  $\delta$  4·52 signal shifted to  $\delta$ 5·65 (d, d 9 Hz), the alcohol function can be assigned to  $C_5$ .

Spin decoupling experiments with (3) established coupling interactions between  $H_5$  and  $H_6$ ; and  $H_6$  with the  $C_{13}$ -vinyl methyl group thus indicating that marginatin (1) is lactonized to  $C_6$ .

The dihydrotiglate moiety present in (3) was hydrogenolyzed by the catalytis hydrogenation of (3) to yield (5) which was also obtained from (1) directly. Since this reaction with (3) must be activated by the  $C_7$ – $C_{11}$  double bond, the tiglate ester must be at  $C_8$  in marginatin (1). Moreover, the multiplicity of the  $H_8$  signal in (3) suggests that  $C_9$  bears at least two protons; therefore, the double bond associated with the  $C_{10}$  vinyl methyl group can be placed between  $C_1$  and  $C_{10}$ . However, since the multiplicity of  $H_8$  in the PMR spectrum of (3) might be due to sterioisomers formed during hydrogenation, (6) was prepared by a controlled catalytis hydrogenation. An analysis of the PMR spectrum of (6), dihydrodeacetoxymarginatin, provided additional support for the presence of the  $C_1$ – $C_{10}$  double bond. The spectrum of (6) exhibited a doublet at  $\delta$  2-01 (3H, J 2 Hz), assigned to the  $C_{13}$  vinyl methyl group; another doublet at  $\delta$  2-70 (1H, J 10), assigned to  $H_5$ ; a doublet of quartets at  $\delta$  4-75 (1H, J 10, 2), assigned to  $H_6$ ; a triplet at  $\delta$  5-10 (1H, J 6), assigned to  $H_8$ ; and a broad triplet at  $\delta$  5-45 (1H, J 7), assigned to  $H_1$ . The multiplicity of the latter signal requires that the vinyl proton  $H_1$  be adjacent to a methylene group, in accord with structure (1) for marginatin.

Comparison of the  $^{13}$ C-NMR spectrum for marginatin with those obtained for glaucolide-A (and B),\* whose total structure was established by X-ray crystallography,<sup>4</sup> indicate that the stereochemistry at the  $C_4$ ,  $C_5$ ,  $C_6$  and  $C_8$  positions in marginatin are the same as those in glaucolide-A and B (2a, b) respectively. No direct data are available to indicate the stereochemistry about the  $C_1$ - $C_{10}$  double bond, although most, but not all, germacranolides containing this function have a *trans* configuration for this double bond.

It is of interest to note that marginatin (1) has a simpler structure than does glaucolide-A (2a) and B (2b) and since (1) dominates in what are considered to be recently evolved species, <sup>3,6</sup> it appears that in these taxa and for these compounds loss mutation is an advanced evolutionary character.

<sup>\*</sup> The <sup>13</sup>C-NMR data will be described in a later paper: Bhacca, N. S., Wehrli, F., Mabry, T. J. and Padolina, W. G., in preparation.

<sup>5 (</sup>a) GEISSMAN, T. A. and GRIFFIN, T. S. (1971) Revista Lationoamer de Química 2, 81; (b) FRAZER, R. R. (1960) Can. J. Chem. 38, 549.

<sup>&</sup>lt;sup>6</sup> JONES, JR., S. B., private communication.

TABLE 1. PMR DATA FOR

Compound	$H_5$	$\mathbf{H}_{6}$	$H_8$	H <sub>13</sub>
(1)†	2·83 (d, 1H, J 8)	4·98 (d. 1H. J 9)	5·20 (t. 1H, J 7)	4·92 (brd s. 2H)
(3)†	2·58 (d, 1H, J 10)	4·88 (dq. 1H. J 10, 2)	5·40 (dd, 1H, J 11, 3)	VM 2·03 (d. 3H, J 2)
( <b>4</b> a) <sup>+</sup>	4·52 (d, 1H, J 9)	5·12 (dq. 1H. J 9, 2)	6·00-5·30 (c. 2H) 5·82-5·10 (c. 2H)	VM 1·93 (d. 6H, J 2) VM 1·95 (d. 6H, J 2)
(4b) <sup>+</sup>	5·65 (d. 1H. J 9)	5·22 (brd, d. 1H. J 9)		
(5)†	2·59 (d. 1H, J 10)	4·57 (brd, d, 1H, J 10)		VM 1·87 (d. 3H. J 1·5)
(6);	2·70 (d. 1H. J 10)	4·75 (dg. 1H, J 10, 2)	5·10 (t, 1H, J 6)	VM 2:01 (d, 3H, J/2)
1† + Pr(fod) <sub>3</sub>	Shifted upfield	4·05 (d. 1H, J 10)	4·7() (dd, 1H. J 10. 4)	3·55 (brd s. 2H)

<sup>\*</sup> Spectra were determined on a Varian A60 spectrometer. Chemical shifts are in  $\delta$  units (ppm) relative to tetramethylsilane as internal standard. Parentheses contain signal multiplicity, number of protons, and coupling constant. J in Hz. Signal multiplicity is designated by the following symbols: s = singlet. d = doublet. t = triplet, dq = doublet composed of two quartets, c = complex. For 3, 4a, 4b and 5 the spectral data are for mixtures of stereoisomers formed during hydrogenation.

## EXPERIMENTAL

Spectra were measured on Varian A60 and HA100 (NMR); Beckman IR5A (IR); CEC110 (MS. 70eV, direct insertion); and Perkin-Elmer 141 polarimeter (specific rotation).

Marginatin (1). Dried and ground leaves (220 g) of Vernonia marginata\* were extracted with CHCl<sub>3</sub> and worked-up in the usual way to yield 3·4 g of crude syrup. Pure marginatin (1) was obtained either by chromatography over a column of silica gel ( $C_6H_6$ : EtOAc, 3:1) or trituration of the crude syrup with  $C_6H_6$ : Et<sub>2</sub>O, m.p.  $104-105^\circ$ ; [ $\alpha$ ]<sub>D</sub><sup>2.5</sup>  $-6\cdot0^\circ$ ; UV (EtOH)  $\lambda_{max} = 215$  nm ( $\epsilon$  25 600): IR 1760, 1710, 1615, 1236 cm<sup>-1</sup>; PMR data are in Table 1. (Anal. Calcd for  $C_{22}H_{28}O_7$ : C, 65·35; H, 6·94; O, 27·72. Found: C, 65·56; H, 6·73; O, 27·40°<sub>6</sub>.)

Hydrogenation of marginatin (1) to tetrahydrodesacetoxymarginatin (3). A solution of 500 mg of 1 in 100 ml of EtOAc was hydrogenated under NTP with 500 mg prereduced  $5^{\circ}_{\circ}$  Pd-C catalyst. The reaction was stopped after the uptake of 3 equivalents of  $H_2$ . The soln was cone in vacuo and purified by preparative TLC to yield, after crystallization from MeOH, 300 mg of fine needles of a mixture of isomers of tetrahydrodesacetoxymarginatin (3): m.p.  $83-85^{\circ}$ ;  $[z]_D^{25} - 87\cdot8^{\circ}$ ; UV (EtOH)  $\lambda_{\text{max}} = 210$  nm ( $\epsilon$  15 800); IR 2950, 1750, 1720, 1205 cm<sup>-1</sup>; PMR data in Table 1. (Anal. Calcd for  $C_{20}H_{34}O_5$ ; (M  $^{\circ}$  + 1), 351·2171. Found: (M  $^{\circ}$  + 1), 351·2180: A weak M  $^{+}$  peak was observed.)

Isomerization of tetrahydrodesacetoxymarginatin (3) to the allylic alcohol 4a. 200 mg of (2) in 4 ml MeOH was added to 0.7 ml of conc HCl and the mixture reacted overnight. The solvents were removed under vacuum and the products separated by preparative TLC ( $C_6H_6$  Et<sub>2</sub>O. 1:2) to yield 80 mg of an oily product (4) (IR 3500, 2950, 1770, 1710 cm<sup>-1</sup>; PMR data in Table 1) characterized as the acetate 4b which also isolated as an oil:  $[\alpha]_D^{1.5} - 62\cdot0^\circ$ ; UV (EtOH)  $\lambda_{max} = 210$  nm ( $\epsilon$  12 200); IR 2950, 1700, 1710, 1240 cm<sup>-1</sup>; PMR data are in Table 1. (Anal. Calcd for  $C_{22}H_{32}O_6$ ; MW, 392·2199. Found: mol wt. (MS), 392·2194.)

Hydrogenation of marginatin (1) to the epoxy lactone (5). A soln of 100 mg of 1 in 100 ml of EtOH was hydrogenated NTP with 100 mg of prereduced PtO<sub>2</sub>. The reaction was stopped after the uptake of 4 equivalents H<sub>2</sub> and after cone, preparative TLC (hexane: Et<sub>2</sub>O, 1:1) and recrystallization from petrol. 70 mg of fine needles of

<sup>\*</sup> Information on the collection sites of V. marginata, V. fasiciculata and V. arkansana and the deposition of vouchers are recorded elsewhere. 1.2

<sup>&</sup>lt;sup>7</sup> Mabry, T. J., Miller, H. E., Kagan, H. B. and Renold, W. (1966) Tetrahedron 22, 1139.

## MARGINATIN AND ITS DERIVATIVES\*

C <sub>4</sub> -Me	C <sub>10</sub> -Me	OAc-Me	Miscellaneous
1.40	1.95–1.66	2.02	Tiglate vinyl methyl 1.95–1.66 (c, 9H)
(s, 3H)	(c, 9H)	(s, 3H)	tiglate vinyl-H 6·90 (c, 1H) C <sub>1</sub> -vinyl proton 5·55 (brd t, 1H, J 7)
1.62	1.05-0.65	_	2-Me-butyrate 1·16 and 1·13 both $(d, J7)$
(s, 3H)	(c, 6H)		integrating for 3H butyrate terminal Me 1·05–0.65 (c, 6H)
Overlapping	1.03	_	2-Me-butyrate 1.07 (d, 3H, J 7)
$W/C_{13}$ - $VM$	(d, 3H, J7)		butyrate terminal-Me 0.89 (dd, 3H, J 6, 2) C <sub>3</sub> -vinyl proton 6.00–5.30 (c, 2H)
			$C_5$ -OH 6·00–5·00 (brd s, 1H) lost w/ $D_2$ O
Overlapping	1.08	2.06	2-Me-butyrate 1.08 (d, 3H, J 7);
$W/C_{13}$ -VM	(d, 3H, J 7)	(s, 3H)	butyrate terminal-Me $0.89$ (dd, 3H, J 6, 2) C <sub>3</sub> -vinyl proton $5.82-5.10$ (c, 2H)
1.55	0.92		
(s, 3H)	(brd s, 3H)		
1.39	1.76		2-Me-butyrate 1·13 and 1·10 both (d, J 7)
(s, 3H)	(d, 3H, J 1)		integrating for 3H butyrate terminal-Me 0.93 (dd, 1H, J 6, 3) C <sub>1</sub> -vinyl proton 5.45 (brd t, 1H, J 7)
1.01/1.16	1.90-1.40	1.01/1.16	Tiglate vinyl methyl $1.90-1.40$ (c, 9H)
(s, 3H)§	(c, 9H)	(s, 3H)§	tiglate vinyl-H 6·67 $(q, 1H, J, 8)$
(0, 511)8	(0, 711)	(0, 511)8	$C_1$ -vinyl proton 5·18 ( $t$ , 1H, $J$ 8)

<sup>†</sup> In chloroform-d.

the epoxy lactone (5) were obtained: m.p.  $138-141^{\circ}$ ;  $[\alpha]_D^{25} - 6\cdot 2^{\circ}$ ; UV (EtOH)  $\lambda_{max} = 212 \text{ nm}$  ( $\epsilon 12000$ ); IR 2950, 1750 cm<sup>-1</sup>; PMR data are in Table 1. (Anal. Calcd for  $C_{15}H_{22}O_3$ : MW, 250·1569. Found: MW (MS), 250·1570.) Hydrogenation of tetrahydrodesacetoxymarginatin (3) to the epoxy lactone (5). A soln of 100 mg of (3) in 20 ml EtOH was hydrogenated at NTP with 100 mg of prereduced PtO2. The reaction was stopped after the absorption of 1 equivalent H<sub>2</sub> and worked-up as the catalyst was filtered off. The filtrate was conc under H<sub>2</sub>O pump vacuum to yield 40 mg of the epoxy lactone 5 (identical m.p. and IR and PMR spectra).

Hydrogenation of marginatin (1) to dihydrodesacetoxymarginatin (6). A soln of 500 mg of marginatin (1) in 500 ml EtOAc was hydrogenated at NTP with 500 mg prereduced 5% Pd-C. The reaction was stopped after the uptake of 2 equivalents of  $H_2$  and worked-up. The catalyst was as before, giving 250 mg of an oily mixture of isomers of 6:  $[\alpha]_D^{2.5}$  - 37·0° UV (EtOH)  $\lambda_{max} = 210$  nm ( $\epsilon$  13 500); IR 2950, 1750, 1710 cm<sup>-1</sup>; PMR data are in Table 1. (Anal. Calcd for  $C_{20}H_{28}O_5$ : MW, 348·1937. Found: MW (MS), 348·1936.)

Acknowledgements—The work was supported by grants from the Robert A. Welch Foundation (Grants F-130 and F-329), and the National Science Foundation (Grants GB-29576X and GB-27152). We thank Dr. S. B. Jones, Jr., for collections and identification of the plant material.

<sup>‡</sup> In acetone-d<sub>6</sub>.

<sup>§</sup> Denotes multiplicity and integration for each signal. VM Vinyl methyl.